# A New Triterpenoid Glycoside from Decaisnea Fargesii

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**Abstract:** A new triterpenoid glycoside, named decaisoside F, has been isolated from *Decaisnea* fargesii. Its structure was determined on basis of chemical method, FAB-MS, homonuclear and heteronuclear correlation experiments, including <sup>1</sup>H—<sup>1</sup>H COSY, <sup>1</sup>H—<sup>13</sup>C COSY, TOSCY, HMBC and NOESY techniques to be 3-O- $\beta$ -D- xylopyranosyl (1 $\rightarrow$ 3)- $\alpha$ -L-rhamnopyranosyl (1 $\rightarrow$ 2)- $\alpha$ -L-arabinopyranosyl hederagenin 28-O- $\beta$ -D-xylopyranosyl (1 $\rightarrow$ 4)- $\beta$ -D-glucopyranosyl (1 $\rightarrow$ 6)- $\beta$ -D-glucopyranoside.

#### Keywords: Decaisnea fargesii; triterpenoid glycoside; decaisoside F; 2D-NMR.

*Decaisnea fargesii* Franch. is a traditional Chinese medicinal plant widely distributed in China, used as an anti-rheumatic and antitussive drug for a long time in Chinese folk medicine<sup>1</sup>. Its methanol extract showed antitumour acidity *in vivo* against  $S_{180}$ , Hepa and Ehrlich cells and its phytochemical studies on saponins has reported<sup>2</sup>. As a continuation of the studies on this plant, we present the spectral and chemical evidence of the structure of a new decaisoside F.

The dried stems (2.23 kg) were percolated with 95% EtOH. After removal of the percolates by evaporators, the residue (380 g) was subjected to macto-porous absorption resin D-101 eluting with aq. MeOH gradiently. The 80% MeOH eluate was concentrated to dryness to give a crude glycoside fraction. This fraction was separated by chromatography (silica gel column and RP-8 column), to afford decaisoside F (860 mg).

Decaisoside F **1**, White powder from MeOH, mp. 227-230 °C (dec.),  $[\alpha]_D^{23} - 18.9$  (MeOH, c 0.009). It responded positively to the Liebermann Burchard test<sup>3</sup>. Its IR spectra showed ester group aborptions (1730-1740 cm<sup>-1</sup>) together with hydroxyl absorptions (3400-3420 cm<sup>-1</sup>), and C=C double bond absorptions (1640-1650 cm<sup>-1</sup>). Its molecular formula was deduced as C<sub>63</sub>H<sub>102</sub>O<sub>29</sub> by means of <sup>13</sup>CNMR, DEPT spectrum and the negative FAB-MS. On acid hydrolysis on TLC<sup>4</sup>, **1** yielded rhamnose, arabinose, glucose and xylose. On acid hydrolysis with 0.2 mol/L HCI in 60% EtOH, **1** afforded hederagenin and prosapogenin **2**. Compound **2** was determined to be hederagenin 3-O- $\beta$ -D-xylopyranosyl (1 $\rightarrow$ 3)- $\alpha$ -L-rhamnopyranosyl (1 $\rightarrow$ 2)- $\alpha$ -L- arabinopyranoside based on the comparison of <sup>1</sup>H, <sup>13</sup>CNMR and FAB-MA as well as the result of monosaccharide

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analysis. The structure of **2** is identical with saponin PG from *Akebia quinata*<sup>5</sup>. The <sup>13</sup>C NMR and CH-COSY spectra of compound **1** showed the presence of six anomeric carbon signals ( $\delta = 95.6$ , 101.2, 104.5, 104.9, 105.0, 107.5 ppm), and six anomeric proton signals ( $\delta = 6.19$ , 6.28, 5.03, 4.92, 5.30, 5.29 ppm) by direct correlation peaks. On comparison of the <sup>13</sup>C NMR spectrum of compound **1** with that of hederagenin, the glycosylation shifts<sup>6</sup>, were observed for C-2, C-3, C-23 and C-28 ( $\delta = 26.3$ , 80.0, 64.0, 176.5 ppm). The signals of anomeric carbon ( $\delta = 95.6$  ppm) and C-28 ( $\delta = 176.5$  ppm) all indicated that **1** was a 28-O-glycosidic linkage. The simultaneous presence of a 3-O-glycosidic linkage was easily seen by the attendant downfield shift ( $\delta = 80.0$  ppm) for C-3. This information showed that **1** was a 3, 28-di-glycoside of hederagenin<sup>7</sup>. The assignment was supported by data from the HMBC spectrum in which the anomeric proton ( $\delta = 5.03$  ppm) showed cross-peak to C-3 ( $\delta = 80.0$  ppm), and anomeric proton ( $\delta = 6.19$  ppm) showed cross-peak to C-28 ( $\delta = 176.5$  ppm).





In the negative FAB-MS of **1**, besides the quasimolecular ion peak, fragment ion peaks at m/z 1337 [M-H]<sup>-</sup>, 1205 [M-Xyl-H]<sup>-</sup>, 881 [M-2Glc-Xyl-H]<sup>-</sup>,749 [M-2Glc-2Xyl-H]<sup>-</sup> were observed. The HMBC spectrum gives the correlation peaks of the terminal xylose H-1 ( $\delta = 5.03$  ppm) to the inner glucose C-4 ( $\delta = 80.7$  ppm) and the inner glucose H-1 ( $\delta = 4.92$  ppm) to inner-28-O-glucose C-6 ( $\delta = 69.5$  ppm); the terminal xylose H-1 ( $\delta = 5.29$  ppm) to the rhamnose C-3 ( $\delta = 82.8$  ppm) and the rhamnose H-1 ( $\delta = 6.28$  ppm) to arabinose C-2 ( $\delta = 74.4$  ppm). The other proton and carbon resonance in the middle of rings were identified by the correlation with anomeric, methylene and methyl protons using COSY, TOCSY, CH-COSY and NOESY spectra (see **Table 1** and **2**). Thus, the structure of **1** was established to be 3-O-  $\beta$  -D- xylopyranosyl (1  $\rightarrow$  3)-  $\alpha$  -L-rhamnopyranosyl (1 $\rightarrow$ 2)- $\alpha$ -L-arabinopyranosyl hederagenin 28-O- $\beta$ -D-xylopyranosyl (1 $\rightarrow$ 4)- $\beta$ -D-glucopyranosyl (1 $\rightarrow$ 6)- $\beta$ -D-glucopyranoside.

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С		Н	С		Н	
1	39.0	1.03α, 1.50β	16	23.8	1.87α, 1.98β	
2	26.3	1.98α, 2.17β	17	47.0		
3	80.0	4.22α	18	41.6	3.13	
4	43.5		19	46.2	1.15α, 1.64β	
5	48.1	1.72 <sub>α</sub>	20	30.7		
6	18.1	1.65a, 1.65b	21	33.9	1.06α, 1.23β	
7	32.5	1.69α, 1.82β	22	32.7	1.21α, 1.55β	
8	38.9		23	64.0	$3.86\alpha, 4.24\beta$	
9	47.6	1.72 <sub>α</sub>	24	14.1	1.10	
10	36.8		25	16.1	0.94	
11	23.3	1.87a, 1.87b	26	17.5	1.07	
12	122.9	5.37	27	26.0	1.15	
13	144.1		28	176.5		
14	42.1		29	33.0	0.82	
15	28.8	1.06α, 2.24β	30	23.6	0.83	

**Table 1** <sup>13</sup>CNMR data for the triterpene moiety of decaisoside F in pyridine-d<sub>5</sub> ( $\delta$  ppm)

Table 2.	<sup>1</sup> HNMR and	<sup>13</sup> CNMR d	lata for the	saccharides	of decaisosi	ide F in p	oyridine-d <sub>5</sub> (	(δ ppm)

		C		TT	
Residue		104.5	5.02		
α-ara p	1	104.5	5.03	(d, J=5.5HZ)	
	2	/4./	4.52		
	3	/5.5	4.00		
	4	69.5	4.06		
1	5	66.0	$5.08\alpha, 4.18\beta$		
α-rha p	1	101.2	6.28	(s)	
	2	71.9	4.86		
	3	82.8	4.72		
	4	72.9	4.42		
	5	69.5	4.66		
	6	18.3	1.55		
β-xyl p	1	107.5	5.29	(d, J=7.5Hz)	
	2	74.9	3.94		
	3	78.2	4.07		
	4	70.9	4.19		
	5	67.2	3.68α, 4.16β		
β-glc p	1	95.6	6.19	(d, J=8.5Hz)	
	2	73.8	4.08		
	3	76.2	4.15		
	4	70.7	4.32		
	5	77.7	4.05		
	6	69.5	4.08α, 4.26β		
β-glc p	1	104.9	4.92	(d, J=7.5Hz)	
	2	74.9	3.98		
	3	76.1	4.15		
	4	80.7	4.20		
	5	76.4	3.76		
	6	61.6	4.35α, 4.45β		
β-xyl p	1	105.0	5.30	(d, J=8.0Hz)	
,	2	74.8	3.94		
	3	78.0	4.08		
	4	70.8	4.12		
	5	67.2	3.61α, 4.17β		

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